

Notes

**A Convenient Synthesis of
(±)-*talo*-Quercitol (1-Deoxy-*neo*-Inositol)
and (±)-*vibo*-Quercitol
(1-Deoxy-*myo*-Inositol) via Ene Reaction of
Singlet Oxygen**

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The polyhydroxy cyclohexanes have been an interest to those concerned with carbohydrates¹. The first known cyclohexanepentol was a dextrorotatory cyclitol obtained from the acorns of *Quercus* species (oaks)², hence the name (+)-*proto*-quercitol. "Quercitol" has been used as a generic term for cyclohexanepentols. Ten diastereoisomers are predicted for the quercitols. All 10 are known and only the (+)-*proto*,² (–)-*proto*,³ and (–)-*vibo*⁴ stereoisomers have been found in nature. Methods which have been used for quercitol synthesis include reduction (hydrogenation) of inososes, inosos oximes, or deoxyinososes; hydrogenation of bromoquercitols; reduction of anhydroinositols; and transformation of conduritols.⁵ Meanwhile, synthesis of *talo*-quercitol and *vibo*-quercitol was described by McCasland⁶ and Nakajima⁷ starting from haloinositols and conduritols-E, respectively. In all previously reported syntheses, starting materials have been natural products or compounds which required many steps to synthesize. Recently we described a simple route leading to the synthesis of quercitols where we applied for the first time a singlet oxygen ene reaction combined with the singlet oxygen [2 + 4] addition⁸ successively to the synthesis of *proto*-quercitol^{9,10} and

gala-quercitol.¹⁰ Here with we describe a short and efficient synthesis of (±)-*talo*-quercitol and (±)-*vibo*-quercitol starting from commercial 1,3-cyclohexadiene where we used again ene reaction of singlet oxygen.¹¹

Results and Discussion

1,4-Cyclohexadiene **1** was used as the starting material for the synthesis of *talo*-quercitol. OsO₄-catalyzed NMO oxidation¹² of **1** afforded diol **2**. The resulting *cis*-diol **2** was protected by ketal formation with 1,2-dimethoxypropane. Tetraphenylporphyrin-sensitized photooxygenation of ketal **3** in methylene chloride at room temperature gave the hydroperoxide **4** in high yield, via ene reaction of singlet oxygen, as the sole product (Scheme 1). The peroxide linkage is highly susceptible to reductive cleavage by a variety of reductants.¹³ Selective reduction of peroxide linkage was performed with thio-urea under very mild conditions to give the alcohol **5a**. Since the only oxygen–oxygen bond breaks in this reaction, it preserves the configuration at the peroxide-bonded carbon atom. For further structural proof, **5a** was converted to the corresponding acetate **5b** which has been fully characterized on the basis of the spectroscopic data.

Proton and carbon NMR studies of **5b** indicated the formation of an unsymmetrical compound as expected. The proton adjacent to the acetoxy group appears as a multiplet at 5.28 ppm, the alkoxy protons at 4.32 ppm as a multiplet, and the methylene protons appear as an AB system at 2.31 and 1.65 ppm. An 11-line ¹³C NMR spectrum is also in agreement with the proposed constitution. However, on the basis of NMR data alone we were not able to assign the correct configuration of the acetoxy group. The configuration of NOE effects. Irradiation at the resonance signals of alkoxy protons at δ = 4.32 caused signal enhancement at the resonances of the adjacent double bond proton, methylenic protons, and especially the methyl resonance (acetyl group) at δ = 1.92. These observations clearly indicate the anti-configuration of acetoxy group in **5b**. Singlet oxygen is quite sensitive to steric consideration and approaches the substrate predominantly, if not exclusively, from the less congested side.¹⁴ The pronounced stereoselectivity of this

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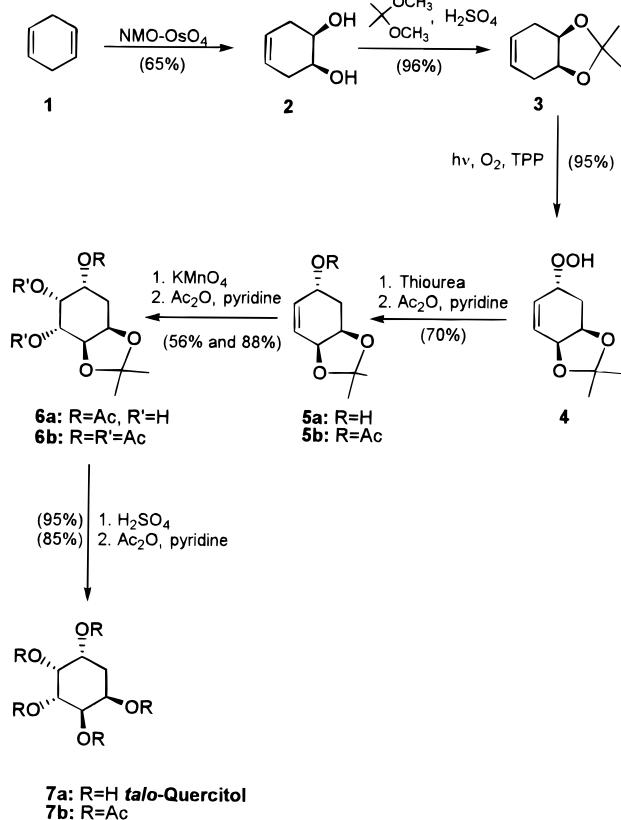
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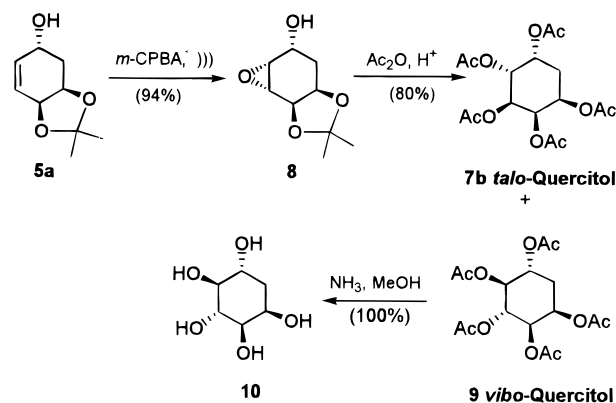
Scheme 1



ene reaction is rationalized by the strong steric effect between the incoming singlet oxygen and bulky methyl groups. In the next step, the double bond has to be oxidized in a *cis* fashion. The oxidation of olefins with permanganate is not commonly used as a preparative method because of its typically low selectivity. However, we chose to examine the oxidation of acetate **5b** with permanganate with the intention of introducing the two hydroxyl groups in a *cis* configuration to the double bond to complete the synthesis of *talo*-quercitol. Treatment of **5b** with KMnO_4 (-5°C) gave to our surprise only one diol **6a** in an isolated yield of 56%. Careful NMR studies did not reveal the formation of any trace of the other diastereomer. For characterization of the product, **6a** was converted to the corresponding triacetate **6b**, which upon hydrolysis gave *talo*-quercitol **7a** (Scheme 1). For further characterization of **7a** we prepared the corresponding pentaacetate in 85% yield. The spectral data of **7a** and **7b** were identical with those reported in the literature.⁶

For the synthesis of *vibo*-quercitol **10**, one should introduce two hydroxyl group to the double bond in **5** in a *trans* fashion (Scheme 2). Therefore, we have submitted **5b** to epoxidation reaction¹⁵ with *m*-chloroperbenzoic acid in an ultrasonic bath and obtained the epoxide in 94% yield as the sole product **8**, whose exact configuration was determined on the basis of NMR spectral data and NOE experiments. The directive effect of the hydroxyl group has been suggested to arise because of hydrogen bonding between the hydroxyl group and the attacking

Scheme 2



peracid.¹⁶ The isolated epoxide **8** was submitted to acid-catalyzed ring-opening reaction in acetic anhydride and we have isolated a mixture consisting of *talo*-pentaacetates **7b** and *vibo*-pentaacetate **9**¹⁵ in a ratio of 4:1. The resulting mixture has been separated on a silica gel column. Ammonolysis of **7b** and **9** furnished the corresponding quercitols.

Experimental Section

General Procedure. Melting points were determined on a melting apparatus. Infrared spectra were obtained from KBr pellets on an infrared spectrophotometer. ^1H NMR spectra were recorded on 200 MHz spectrometer and reported in δ units with SiMe_4 as internal standard. All column chromatography was performed on silica gel (60 mesh).

4-Cyclohexene-*cis*-1,2-diol (2). The reported procedure¹² was used for the synthesis of 4-cyclohexene-*cis*-1,2-diol **2** in 65% yield: mp $69-70^\circ\text{C}$, colorless crystals from ethyl acetate:hexane (3:1); ^1H NMR (200 MHz, CDCl_3) δ 5.55 (m, 2H), 3.91 (m, 2H), 2.28 (m, 4H); ^{13}C NMR (50 MHz, CDCl_3) δ 125.70, 70.90, 32.93.

(3a,R,7a,S)-2,2-Dimethyl-3a,4,7,7a-tetrahydrobenzo-1,3-dioxole (3). The described ketalization procedure in the literature^{8a,17} was applied to the synthesis of **3** (colorless liquid, 96%); ^1H NMR (200 MHz, CDCl_3) δ 5.80 (m, 2H), 4.35 (m, 2H), 2.25 (m, 4H), 1.41 (s, 3H), 1.32 (s, 3H); ^{13}C NMR (CDCl_3 , 50 MHz) δ 126.00, 107.97, 73.56, 28.78, 27.49, 25.35.

(1a,4b,6a)-8,8-Dimethyl-7,9-dioxabicyclo[4.3.0^{1,6}]non-2-en-4-yl hydroperoxide 4. To a stirred solution of ketal **3** (1.0 g, 6.4 mmol) in 150 mL of CH_2Cl_2 was added 20 mg of tetraphenylporphyrin (TPP). The resulting mixture was irradiated with a tungsten halogen projection lamp (500 W) while oxygen was being passed through the solution and the mixture was stirred at room temperature for 12 h. Evaporation of the solvent (30°C , 20 mmHg) and chromatography of the residue on a Florisil column (30 g) eluting with *n*-hexane/ CH_2Cl_2 (7:4) gave pure hydroperoxide **4** (1.15 g, 95%, colorless liquid): ^1H NMR (200 MHz, CDCl_3) δ 8.15 (s, 1H), 6.06 (dm, A part of AB system, $J = 10.4$ Hz, 1H), 5.95 (dm, B part of AB system, $J = 10.4$ Hz, 1H), 4.70 (m, 1H), 4.68 (m, 2H), 2.45 (dt, A part of AB system, $J = 13.6$, 3.9 Hz, 1H), 1.95 (ddd, B part of AB system, $J = 13.6$, 8.8, 2.8 Hz, 1H), 1.37 (s, 3H), 1.36 (s, 3H); ^{13}C NMR (CDCl_3 , 50 MHz) δ 129.72, 129.46, 109.29, 77.21, 72.85, 71.85, 30.15, 28.24, 26.82; IR (KBr, cm^{-1}) 3462, 3387, 3004, 2953, 1651, 1472, 1395, 1242, 1165, 1063, 859.

(1a,4b,6a)-8,8-Dimethyl-4-hydroxy-7,9-dioxabicyclo[4.3.0^{1,6}]non-2-ene 5a. To a magnetically stirred slurry of 0.82 g (10.7 mmol) of thiourea in 25 mL of methanol was added a solution of 1.00 g (5.37 mmol) of *anti*-hydroperoxide **4** in 50 mL of methanol at 25°C . After complete of addition (ca. 10 min),

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the mixture was stirred for 1 h, the solid was removed by filtration, methanol was evaporated (35 °C, 20 mmHg), and the residue purified by column chromatography on silica gel (10 g) eluting with chloroform afforded pure **5a** (0.82 g, 90%, colorless liquid): ¹H NMR (200 MHz, CDCl₃) δ 5.86 (dm, A part of AB system, *J* = 11.4 Hz, 1H), 5.65 (ddd, B part of AB system, *J* = 11.4, 4.3, 2.4 Hz, 1H), 4.45 (m, 3H), 2.44 (ddm, A part of AB system, *J* = 13.8, 3.7 Hz, 1H), 1.95 (ddd, B part of AB system, *J* = 13.8, 9.5, 2.4 Hz, 1H), 1.32 (s, 6H); ¹³C NMR (CDCl₃, 50 MHz) δ 135.34, 129.10, 109.30, 73.40, 71.91, 63.82, 35.83, 28.12, 26.35; IR (KBr, cm⁻¹) 3438, 3004, 2953, 2902, 1472, 1395, 1242, 1089, 1038, 885. Anal. Calcd for C₉H₁₄O₃: C, 63.51; H, 8.29. Found: C, 63.30; H, 8.12.

(1a,4b,6a)-8,8-Dimethyl-4-acetoxy-7,9-dioxo-bicyclo[4.3.0^{1,6}]-non-2-ene 5b. To a magnetically stirred solution of alcohol **5a** (0.6 g, 3.50 mmol) in 3 mL of pyridine was added acetic anhydride (0.428 g, 4.2 mmol). The reaction mixture was stirred at room temperature for 8 h and cooled to 0 °C. After addition of 100 mL of water, the water phase was extracted with ether (3 × 50 mL). The combined organic extracts were washed with NaHCO₃ solution (10 mL) and water (5 mL) and then dried (Na₂SO₄). Removal of the solvent under reduced pressure gave acetate **5b** (0.59 g, 78%, colorless liquid): ¹H NMR (200 MHz, CDCl₃) δ 5.71 (dm, A part of AB system, *J* = 10.8 Hz, 1H), 5.65 (dm, B part of AB system, *J* = 10.8 Hz, 1H), 5.28 (m, 1H), 4.32 (m, 2H), 2.31 (ddt, A part of AB system, *J* = 13.8, 5.5, 1.1 Hz, 1H), 1.65 (ddd, B part of AB system, *J* = 13.8, 9.1, 2.8 Hz, 1H), 1.92 (s, 3H), 1.23 (s, 3H), 1.21 (s, 3H); ¹³C NMR (CDCl₃, 50 MHz) δ 171.15, 130.05, 129.10, 109.12, 72.22, 71.83, 66.52, 31.80, 28.12, 27.82, 22.05; IR (KBr, cm⁻¹) 2990, 1740, 1370, 1240, 1040. Anal. Calcd for C₁₁H₁₆O₄: C, 62.25; H, 7.60. Found: C, 61.93; H, 7.46.

(1a,2b,3b,4b,6a)-8,8-Dimethyl-2,3,4-triacetoxy-7,9-dioxo-bicyclo[4.3.0^{1,6}]-nonane 6b. To a magnetically stirred ethanol solution (100 mL) of alkene **5b** (2.5 g, 11.79 mmol) was added a solution of KMnO₄ (1.86 g, 11.79 mmol) and MgSO₄ (1.41 g, 11.79 mmol) in water (40 mL) at -5 °C during 7 h. After the addition was completed, the reaction mixture was stirred for an additional 15 h at the given temperature and then filtered. The precipitate was washed several times with hot water. The combined filtrates were concentrated to 20 mL by rotoevaporation. The aqueous solution was extracted with ethyl acetate (3 × 30 mL), and the extracts were dried (Na₂SO₄). Evaporation of the solvent gave diol **6a** (1.630 g, 56%), which was submitted to acetylation as described above to give **6b** (1.92 g, 88%, colorless liquid): ¹H NMR (200 MHz, CDCl₃) δ 5.41 (m, 1H), 5.15 (ddd, *J* = 11.9, 5.5, 1.75 Hz, 1H), 4.95 (dd, *J* = 8.4, 2.5 Hz, 1H), 4.35 (m, 1H), 4.16 (dd, *J* = 11.9, 8.4 Hz, 1H), 2.30 (dd, A part of AB system, *J* = 14.2, 5.0 Hz, 1H), 2.13 (dd, B part of AB system, *J* = 14.2, 4.8 Hz, 1H), 2.06 (s, 3H), 1.99 (s, 3H), 1.94 (s, 3H), 1.44 (s, 3H), 1.29 (s, 3H); ¹³C NMR (CDCl₃, 50 MHz) δ 170.42, 170.37, 170.16, 109.92, 75.48, 73.09, 72.41, 70.92, 67.07, 28.47, 27.04, 26.54, 21.27; IR (KBr, cm⁻¹) 2993, 1740, 1435, 1360, 1250, 1060. Anal. Calcd for C₁₅H₂₂O₈: C, 54.54; H, 6.71. Found: C, 54.81; H, 6.54.

(1a,2a,3a,4b,5b)-Cyclohexanepentol [(±)-*talo*-Quercitol] 7a. Ketal triacetate **6b** (1.5 g, 4.5 mmol) was dissolved in 5 mL of 1 N H₂SO₄ and the resulting mixture was stirred at room temperature for 3 h. The acid was neutralized with BaCO₃. Filtration of the precipitate and evaporation of the solvent under reduced pressure gave *talo*-quercitol **7a** (0.7 g, 95%): mp 245–247 °C (lit.^{6a} mp 246–248 °C) from EtOH; ¹H NMR (200 MHz, D₂O) δ 4.0 (m, 3H), 3.85 (m, 2H), 1.80 (dd, *J* = 9.7, 3.1 Hz, 2H); ¹³C NMR (50 MHz, D₂O) δ 75.72, 73.38, 72.69, 70.82, 68.75, 35.17; IR (KBr, cm⁻¹) 3395, 2905, 2500, 1350, 1090, 1000, 845.

(1a,2a,3a,4b,5b)-Pentaacetoxycyclohexane [(±)-*talo*-Quercitol Pentaacetate] 7b. *talo*-Quercitol **7a** was submitted to acetylation as described above to give *talo*-quercitol pentaacetate **7b** (colorless solid, 85%): mp 169–171 °C (lit. mp 182–183 °C,⁶ 169.5–171.5 °C⁷) from EtOH; ¹H NMR (200 MHz, CDCl₃) δ 5.61 (m, 1H), 5.51 (q, *J* = 3.2 Hz, 1H), 5.27 (dd, A part of AB system,

J = 10.8, 2.72 Hz, 1H), 5.21 (dd, B part of AB system, *J* = 10.8, 3.2 Hz, 1H), 5.25 (m, 1H), 2.14 (s, 3H), 2.13 (s, 3H), 2.04 (m, 2H), 2.00 (s, 9H); ¹³C NMR (CDCl₃, 50 MHz) δ 170.72, 170.52, 170.45, 170.20, 70.07, 69.51, 68.20, 67.31, 66.56, 29.36, 21.45, 21.28, 21.19, 21.13; IR (KBr, cm⁻¹) 2995, 1745, 1370, 1230, 1045.

(1a,2b,4b,5b,7a)-9,9-Dimethyl-5-hydroxy-3,8,10-trioxatricyclo[5.3.0^{2,4}.0^{1,7}]decane 8. To a solution of alkene **5a** (1 g, 5.8 mmol) in 5 mL of CH₂Cl₂ were added 2 g of Na₂CO₃ and 1.7 g (6.96 mmol) of *m*-chloroperbenzoic acid (*m*-CPBA). The mixture was sonicated in an ultrasonic bath (50 kHz) for 3 h. The precipitate was filtered and washed with 50 mL of CH₂Cl₂. The combined CH₂Cl₂ solutions were filtered from a silica gel column (5 g) eluting with CH₂Cl₂. Evaporation of the solvent gave **8** (1.01 g, 94%): mp 60–61 °C, colorless solid from CH₂Cl₂; ¹H NMR (200 MHz, CDCl₃) δ 4.31 (m, 3H), 3.39 (br.d, A part of AB system, *J* = 4.00 Hz, 1H), 3.20 (dt, B part of AB system, *J* = 4.0, 1.1 Hz, 1H), 2.15 (ddd, *J* = 14.3, 5.6, 2.4 Hz, 1H), 1.61 (ddd, *J* = 14.3, 10.8, 1.4 Hz, 1H), 1.40 (s, 3H), 1.33 (s, 3H); ¹³C NMR (CDCl₃, 50 MHz) δ 109.66, 73.93, 70.35, 64.39, 56.38, 55.72, 27.98, 27.58, 25.72; IR (KBr, cm⁻¹) 3438, 3004, 2978, 1421, 1395, 1268, 1191. Anal. Calcd for C₉H₁₄O₄: C, 63.68; H, 8.25. Found: C, 63.42; H, 8.11.

Acetolysis of Epoxy Ketal 8. Ketal **8** (500 mg, 2.68 mmol) was dissolved in 5 mL of acetic anhydride, to it was added 50 mg of concentrated H₂SO₄, and the resulting mixture was refluxed for 6 h. The mixture was added to 50 mL of water, extracted with ethyl acetate (3 × 50 mL), and dried (Na₂SO₄). Evaporation of the solvent under reduced pressure gave a mixture consisting of *talo*-quercitol pentaacetate **7b** and *vibo*-quercitol pentaacetate **9** in a ratio of 80:20 determined by ¹H NMR (0.8 g, combined yield 80%). Pentaacetates **7b** and **9** were separated by column chromatography using silica gel and eluting with hexane:ethyl acetate (solvent composition was graded from starting 85:15 to 75:25) to give *talo*-quercitol pentaacetate **7b** (0.508 g, 50%) and *vibo*-quercitol pentaacetate **9** (0.103 g, 10%): mp for **9**, 113–114 °C (lit. mp 126 °C,¹⁹ 114 °C,^{6a} 112–113 °C¹⁸) from ethanol; ¹H NMR (200 MHz, CDCl₃) δ 5.46 (dd, *J* = 3.4, 6.5 Hz, 1H), 5.44 (t, *J* = 9.7 Hz, 1H), 5.28 (dd, *J* = 1.8, 4.3 Hz, 1H), 5.15 (q, *J* = 9.7 Hz, 1H), 4.94 (dd, *J* = 3.2, 10.3 Hz, 1H), 2.28 (dt, *J* = 4.3, 8.6 Hz, 1H), 2.14 (s, 3H), 2.01 (s, 6H), 2.00 (s, 3H), 1.98 (s, 3H), 1.75 (m, 1H); ¹³C NMR (50 MHz, CDCl₃) δ 171.79, 171.85, 171.72, 171.79, 171.66, 75.21, 73.43, 71.67, 70.37, 68.80, 32.74, 22.86, 22.76, 22.52; IR (KBr, cm⁻¹) 3489, 3029, 2953, 1753, 1446, 1395, 1242.

(1a,2a,3b,4b,5a)-Pentahydroxycyclohexane [(±)-*vibo*-Quercitol] 10. A 100 mg (0.26 mmol) portion of *vibo*-quercitol pentaacetate **9** was dissolved in 15 mL of methanol. While dry NH₃ was passed through the solution, the mixture was stirred for 2 h at room temperature. Evaporation of methanol and acetamide gave *vibo*-quercitol **10** (43 mg, quantitative): mp 161–162 °C (lit. mp 163 °C,^{6a} 161–163 °C,¹⁶ 159 °C⁷) from EtOH; ¹H NMR (200 MHz, D₂O) δ 4.07 (br q, *J* = 3.0 Hz, 1H), 3.77 (ddd, *J* = 12.0, 9.4, 4.8 Hz, 1H), 3.57 (t, *J* = 9.2 Hz, 1H), 3.50 (dd, *J* = 3.0, 9.2 Hz, 1H), 3.25 (t, *J* = 9.2 Hz, 1H), 2.10 (dt, *J* = 14.1, 3.9 Hz, 1H), 1.56 (ddd, *J* = 14.3, 12.1, 2.5 Hz, 1H); ¹³C NMR (50 MHz, D₂O) δ 81.80, 78.12, 77.11, 72.77, 72.77, 39.44; IR (KBr, cm⁻¹) 3361, 2927, 1421, 1114, 1038, 987, 680.

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